

Reported analgesic and anaesthetic administration to rodents undergoing experimental surgical procedures

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Abstract

A structured literature review was carried out to assess recent trends in the administration of analgesics and anaesthetics to laboratory rats and mice undergoing surgical procedures. The ScienceDirect database was used to systematically identify studies published in peer-reviewed journals over two periods (2000–2001 and 2005–2006), 86 studies from each time period were included in the review. The total number of animals that underwent surgery, species used, type of procedure, anaesthetic regimen and analgesic administration were noted for each study. There was an increase in the reported administration of systemic analgesics from 10% in 2000–2001 to 20% in 2005–2006. Buprenorphine was the most commonly reported analgesic in both periods (2000–2001: 78%, 2005–2006: 35%) and reporting the use of non-steroidal anti-inflammatory drugs increased from 11% to 53%. There was also a change in reported anaesthetic practices, notably a decrease in the use of pentobarbital and an increase in the use of isoflurane and ketamine/xylazine. Although reported administration of analgesics has increased and there has been some refinement in the selection of anaesthetic agents used, the findings of this review suggest that there is still significant scope for improvement with respect to the perioperative care of laboratory rodents.

Keywords: Anaesthesia, analgesia, pain, rodents, pain reduction

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It is widely accepted that when animals are used in experimental procedures, then measures should be taken to minimize pain and distress.^{1–3} Pain and distress in animals undergoing surgical procedures can be alleviated by providing appropriate perioperative care, which includes the administration of effective anaesthetic and analgesic agents. Although there is a general consensus that one should attempt to alleviate the pain that may be associated with experimental surgery in animals and some studies suggest that analgesics are routinely administered,^{4,5} the administration of analgesics to laboratory rodents is often not reported in peer-reviewed journals. In an earlier literature review carried out by our group we found that the administration of a systemic analgesic agent was reported in only 14% of studies published between 2000 and 2002 that involved laboratory rodents undergoing potentially painful experimental procedures.⁶ In this previous study, we concluded that in the majority of cases, analgesics were withheld rather than analgesic administration being under-reported based on follow-up email inquiries to study authors and institutional veterinarians.

A criticism of this earlier study was that the search strategy was unbalanced and therefore certain areas of research, for example transplantation research, could have been

over-represented. In this review, a more structured search strategy was used to identify areas of research where rodents may undergo recovery surgical procedures. Here, 86 studies published between 2000 and 2001 have been identified from a wide range of fields including pharmacology, surgical research, physiology, neuroscience, immunology and cancer research. Eighty-six papers from these fields published between 2005 and 2006 have also been reviewed to examine changes in the perioperative care of laboratory rodents over these 5 years.

Materials and methods

Search strategy

The ScienceDirect online database (www.science-direct.com) was used to identify relevant studies published in English from 2000 to 2001 and from 2005 to 2006. ScienceDirect was accessed between 15 January 2007 and 12 February 2007. There were two stages to the literature search: initially appropriate journals from a range of biomedical fields were identified, then relevant papers within these journals were selected. To identify journals from a variety of scientific disciplines that published potentially

relevant articles, the search terms 'rodent' and 'surgery' were used. Only journals that were available in electronic format at Newcastle University (approximately 10,000 journals) were examined. Titles and abstracts were then screened to identify journals from a range of biomedical fields.

Once potential journals were selected, the ScienceDirect database was again used with the key terms 'rodent' and 'surgery' to search the full text of articles published within the journals. The target for each time period was to find 10 papers from each journal that met inclusion criteria (see below). If 10 papers could not be found using these key terms, the terms 'rat' and 'surgery' and the terms 'mice' and 'surgery' were used. If a single journal contained less than the target of 10 relevant papers, the key terms were used to search multiple journals within a field of research (e.g. immunology).

Where more than 10 appropriate papers in a single journal were identified from the database, each paper was given a number (based on the ScienceDirect display either by date or by relevance) and a random number generator was used to select 10 of these papers. Similarly, when there were less than 10 relevant studies in one period and over 10 in the other time period, a random number generator was used to select an equal number of papers from the time period with more relevant papers.

Inclusion criteria

A paper was eligible for inclusion if it involved the use of rats or mice undergoing experimental surgical procedures under general anaesthesia with a postoperative recovery period of at least 24 h. All papers included in this review had methods that were described in detail. Papers that: (i) described multiple studies on different species of animals; (ii) described fetal surgery; (iii) described neuropathic models of pain and (iv) stated the efficacy of analgesia following surgery as the purpose of the study was excluded. Review articles, abstracts, letters and meta-analyses were also excluded.

Classification

The classification of the severity of the experimental procedure was based on criteria used in our earlier literature review,⁶ also similar to the classification of the pain potential described by the American College of Laboratory Animal Medicine (ACLAM) Analgesic Task Force.⁷ Each paper was classified into one of the five categories: craniotomy, skin incision, laparotomy, thoracotomy or orthopaedic study. Thoracotomies and orthopaedic procedures were considered to be the most potentially painful procedures, laparotomies were considered to be slightly less potentially painful and skin incisions and craniotomies were considered to be the least potentially painful.

The number of animals that underwent surgery in each study ('study size'), anaesthetic and analgesic regimens were noted. Classification of systemic analgesics was based on Lamont and Matthews⁸ where opioids and non-steroidal anti-inflammatory drugs (NSAIDs) were classified

as systemic analgesic agents. The dose rate, duration, frequency and time of analgesic administration were noted when specified. The use of local anaesthetic agents was also noted. Anaesthetic agents were also classified according to whether they contained an analgesic component. Animals anaesthetized with a dissociative anaesthetic agent (ketamine or tiletamine) and/or an α_2 agonist (medetomidine or xylazine) were classified as having 'an anaesthetic regimen with an analgesic component'. Similarly, animals anaesthetized with an anaesthetic combination that included a fentanyl component were also considered to have received an analgesic component in their anaesthetic. Fentanyl was not classified as a systemic analgesic because of its short-acting effect,⁸ and therefore it is typically given as a component of an anaesthetic regimen, rather than as a systemic analgesic in rodents.

Statistics

All statistical analyses were conducted using SPSS software (SPSS 14.0 statistical package for Windows, SPSS Inc, Chicago, IL, USA). Chi-square analyses were used for all comparisons except a Mann-Whitney test was used to compare 'study size'. A value of $P < 0.05$ was considered statistically significant.

Results

One hundred and seventy-two papers, 86 from each time period (2000–2001 and 2005–2006) were selected from 10 journals/fields of research for inclusion in this review (Table 1).

Table 1 Journals included in review classified by species and time period

Journal name/field name	Number of papers in review			
	2000–2001		2005–2006	
Species	Rats	Mice	Rats	Mice
Journal name				
<i>European Journal of Pharmacology</i>	10	0	10	0
<i>International Journal of Radiation Oncology</i>	4	1	3	2
<i>Journal of Hepatology</i>	9	1	7	3
<i>Journal of Orthopaedic Research</i>	9	1	7	3
<i>Neuroscience</i>	9	1	9	1
<i>Peptides</i>	10	0	8	2
<i>Physiology and Behaviour</i>	8	2	9	1
Field name				
Surgery*	10	0	10	0
Pathology [†]	5	1	4	2
Immunology [‡]	3	2	2	3
Total number of papers	77	9	69	17

* *Annals of Thoracic Surgery, European Journal of Cardio-Thoracic Surgery, Journal of Cranio-Maxiofacial Surgery, Journal of Gastrointestinal Surgery, Journal of Pediatric Surgery, Journal of Vascular Surgery, Surgery*

[†] *Cardiovascular Pathology, Experimental and Molecular Pathology, Experimental and Toxicological Pathology, Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology and Endodontology*

[‡] *Cellular Immunology, Developmental and Comparative Immunology, Transplant Immunology*

Table 2 Median number of animals used in study classified by species and time period

Time period	Rats	Mice
2000–2001	32 (8, 186), <i>n</i> = 72	65 (12, 264), <i>n</i> = 9
2005–2006	47 (5, 224), <i>n</i> = 54	40 (12, 300), <i>n</i> = 11

The minimum and maximum 'study size' are indicated in parentheses
n: number of papers that specified 'study size'

Species and number ('study size') of animals

More studies describing procedures involving rats compared with procedures involving mice were included in this review for both time periods (Table 1). The median number of animals that underwent a surgical procedure in each study was 40 with a minimum and maximum 'study size' of 5 and 300. The number of animals used in the study was not reported in 26 papers. Median 'study size' did not vary significantly with either time period or species (Table 2).

Analgesic administration

There was an increase in the reported administration of systemic analgesics from 10% in 2000–2001 to 20% in 2005–2006. When systemic analgesia was not reported, there was an increase in the use of anaesthetic regimens with an analgesic component between the time periods from 28% to 34%. There was a decrease in the use of local anaesthetic agents between the time periods. Five papers from 2000–2001 specified the application of a local anaesthetic to the surgical site (3 rat craniotomies, 1 rat laparotomy and 1 mouse laparotomy), however, none of the papers from 2005 to 2006, reported the use of a local anaesthetic agent. Neither reported systemic analgesic administration nor total analgesic administration varied significantly between time periods. Table 3 shows analgesic administration classified by the species that underwent surgery.

Buprenorphine was the most commonly reported systemic analgesic both in 2000–2001 (78%) and in 2005–2006 (35%) (Table 4). There was a large increase in the

Table 3 Number of papers included in survey specifying analgesic administration classified by species and time period

Analgesic administration	2000–2001		2005–2006	
	Rats <i>n</i> = 77 (%)	Mice <i>n</i> = 9 (%)	Rats <i>n</i> = 69 (%)	Mice <i>n</i> = 17 (%)
Systemic analgesic specified	8 (10.4)	1 (11.1)	15 (21.7)	2 (11.7)
No systemic analgesic, but analgesic component in anaesthetic	19 (24.7)	5 (55.6)	22 (31.9)	7 (41.2)
Use of a local anaesthetic	4 (5.2)	1 (11.1)	0 (0)	0 (0)
No analgesic	46 (58.7)	2 (22.2)	32 (46.4)	8 (47.1)

Percentages of papers specifying analgesic administration are given in parentheses. *n*: number of papers included in survey

Table 4 Reported systemic analgesic administration classified by agent

Class of analgesic	Analgesic	Number of papers specifying systemic analgesic administration			
		2000–2001		2005–2006	
		Rat	Mouse	Rat	Mouse
Opioids	Buprenorphine	6	1	5	1
	Butorphanol	0	0	1	0
	Tramadol	1	0	0	0
Non-steroidal anti-inflammatory drugs (NSAIDs)	Carprofen	0	0	3	0
	Flunixin meglumine	1	0	2	0
	Ketorolac	0	0	1	0
	Metamizol	0	0	1	0
	Paracetamol	0	0	2	0
	Unspecified	0	0	0	1
Total		8	1	15	2

reported use of NSAIDs between the two time periods: only one paper reported the use of a NSAID in 2000–2001 (flunixin meglumine was administered), whereas 53% of papers that specified the use of an analgesic in 2005–2006 used NSAIDs (Table 4).

When administration of a systemic analgesic was reported, the dose rate administered was specified in the majority of papers (78% in 2000–2001 and 76% in 2005–2006) (Table 5). The duration of systemic analgesic administration was specified less frequently (Table 6). In 2000–2001, four papers specified the duration of systemic analgesic administration (for 1–5 days) and three papers from 2005–2006 specified the duration of analgesic administration (for 1–2 days). The frequency of analgesic administration was also infrequently specified. When frequency of analgesic administration was specified, it varied from a single dose to repeat administration every 24 h with a dose interval of every 12 h being the most commonly reported (Table 6). Only three papers in the survey, all published between 2005 and 2006, specified the time of systemic analgesic administration. One paper specified postoperative administration and two papers (1 mouse thoracotomy and 1 rat orthopaedic study) specified preoperative administration of systemic analgesia.

Table 5 Range of reported doses of systemic analgesics when specified

Time period	2000–2001	2005–2006
Systemic analgesic		
Buprenorphine–mice	0.05 mg/kg (<i>n</i> = 1)	0.03 mg/kg (<i>n</i> = 1)
Buprenorphine–rats	0.01–0.05 mg/kg (<i>n</i> = 5)	0.01–0.1 mg/kg (<i>n</i> = 5)
Butorphanol		1 mg/kg (<i>n</i> = 1)
Carprofen		5–10 mg/kg (<i>n</i> = 3)
Flunixin	2 mg/kg (<i>n</i> = 1)	2.5 mg/kg (<i>n</i> = 1)
Ketofen		4.5 mg/kg (<i>n</i> = 1)
Paracetamol		1 g/L in drinking water (<i>n</i> = 1)

All information is for rats except buprenorphine where dose ranges for mice are also included

Table 6 Duration, frequency and timing of analgesic administration (if specified) in rats

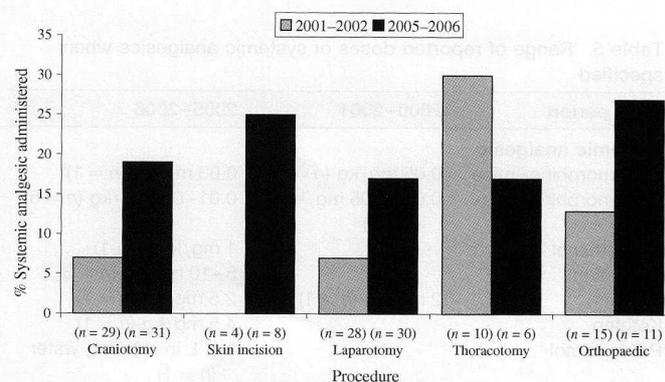
Systemic analgesic agent	Time period	Duration of systemic analgesic administration	Frequency and time analgesic administered, if specified
Buprenorphine	2000–2001	1 day (<i>n</i> = 1), 2 days (<i>n</i> = 1)	q. 12 h (<i>n</i> = 2)
	2005–2006	1 day (<i>n</i> = 1), 2 days (<i>n</i> = 1)	q. 12 h (<i>n</i> = 2)
Butorphanol	2005–2006		1 injection, preoperatively (<i>n</i> = 1)
Carprofen	2005–2006		1 injection, time not specified (<i>n</i> = 1)
Flunixin	2000–2001	2–3 days (<i>n</i> = 1)	q. 24 h (<i>n</i> = 1)
	2005–2006		1 injection, postoperatively (<i>n</i> = 1)
Paracetamol	2005–2006	2 days in drinking water (<i>n</i> = 1)	
Tramadol	2000–2001	5 days in drinking water (<i>n</i> = 1)	

Multimodal analgesia

There were no papers in the study that described the use of more than one systemic analgesic agent or that described the use of a local anaesthetic agent combined with a systemic analgesic or a general anaesthetic regimen with an analgesic component. Ten papers (9 describing procedures in rats, 1 describing a procedure in mice) included in the survey described the use of a systemic analgesic and an anaesthetic regimen with an analgesic component (2 in 2000–2001 and 8 in 2005–2006).

Classification of procedures

The distribution of procedures included in the review varied slightly between the time periods (Figure 1), but the difference was not statistically significant. In the 2000–2001 papers, systemic analgesics were most commonly administered to laboratory rodents undergoing the most potentially painful procedures (thoracotomies and orthopaedic

**Figure 1** Reported use of systemic analgesics classified by category of procedure. *n*: number of papers included in literature review

procedures) compared with less potentially painful procedures (skin incisions and craniotomies). The reported administration of systemic analgesic drugs in the 2005–2006 papers was similar for all procedures and analgesic administration varied from 17% to 27% (Figure 1). The papers included in the survey did not differ significantly between rats and mice when classified according to the potential of the procedure to be painful.

Anaesthetic administration

Pentobarbital was the most commonly used anaesthetic agent in 2000–2001 (Table 7). The use of pentobarbital decreased from 33% in 2000–2001 to 16% in 2005–2006. There was an increase in the use of isoflurane (from 2% to 16%) and in the use of the ketamine/xylazine injectable anaesthetic combination (from 15% to 31%) between the time periods.

Table 7 Reported anaesthetic regimens

	Rats 2000– 2001 <i>n</i> = 77 (%)	Mice 2000– 2001 <i>n</i> = 9 (%)	Rats 2005– 2006 <i>n</i> = 69 (%)	Mice 2005– 2006 <i>n</i> = 17 (%)
Inhaled agent only				
Ether	6 (7.8)		4 (5.8)	1 (5.9)
Halothane	5 (6.5)	1 (11.1)	5 (7.2)	
Isoflurane	2 (2.6)		12 (17.4)	2 (11.8)
Methoxyflurane	2 (2.6)			
	1 (1.3)			
Injectable agent(s) with no analgesic properties				
Chloral hydrate	4 (5.2)		2 (2.9)	
Pentobarbital	26 (33.8)	2 (22.2)	9 (13)	5 (29.4)
Other*	6 (7.8)		2 (2.9)	1 (5.9)
Injectable agent(s) with analgesic properties				
Fentanyl and fluanisone, diazepam and/or medetomidine	1 (1.3)		4 (5.8)	1 (5.9)
Ketamine	2 (2.6)		3 (4.3)	
Ketamine and xylazine	10 (13)	3 (33.3)	22 (31.9)	5 (29.4)
Ketamine and acepromazine, diazepam or pentobarbital	6 (7.8)	1 (11.1)	1 (1.4)	
Other†	2 (2.6)	1 (11.1)	1 (1.4)	1 (5.9)
Combination of injectable and inhaled agents with no analgesic properties‡				
	4 (5.2)	1 (11.1)	4 (5.8)	1 (5.9)

n: number of papers included in the survey. Percentages of papers reporting anaesthetic regimens are given in parentheses. Number of papers included in survey is classified by species and time period

*Methohexital, thiopental, tribromoethanol, chloral hydrate/pentobarbital, pentobarbital/atropine or methohexital/pentobarbital

†Ketamine/xylazine/acepromazine, ketamine/diazepam/atropine or medetomidine/tiletamine/zolazepam

‡Pentobarbital/methoxyflurane, pentobarbital/sevoflurane, chloral hydrate/methoxyflurane, pentobarbital/ether, ether or pentobarbital or chloral hydrate/pentobarbital/isoflurane

Discussion

The reporting of the administration of systemic analgesic drugs to laboratory rodents undergoing surgical procedures is increasing. In 1990–1992, only 3% of studies published in peer-reviewed journals reported analgesic administration.⁶ Here, we report that in 2000–2001 analgesics were administered in 10% of studies and by 2005–2006, analgesics were reported in 20% of studies. Unfortunately however, the majority of papers that describe potentially painful procedures on laboratory rodents still do not report systemic analgesic administration.

Although more studies involving laboratory rats were included in this survey, compared with studies involving laboratory mice (Table 1), this does not reflect the actual use of laboratory rodents as more mice than rats are used in scientific procedures in the UK⁹ and worldwide.¹⁰ Because of their size, rats may be preferable to mice for certain surgical procedures¹¹ and our search strategy may also have resulted in the inclusion of more rat studies compared with mouse studies, because of the large proportion of murine studies that involve the production of genetically-altered animals. In 2006, 47% of all mice used in scientific procedures in Great Britain were genetically modified.⁹ The production of transgenic mice typically involves the use

of standardized surgical procedures that would not be normally described in peer-reviewed journals, for example embryo transfer and vasectomy procedures, which are likely to be painful.^{12,13}

Analgesic administration did not differ significantly between species, however this may have been due to the relatively small sample size of papers describing surgical procedures in mice in this study and therefore a different search strategy would be necessary to identify species differences. Similarly 'study size' did not differ significantly with either time period or species (Table 2), but this may also have been due to the small sample size of murine papers in the survey and the short interval (5 years) separating the two time periods. Previous studies that examined papers over a longer time period have reported a decrease in the number of animals used per published paper (for example Carlsson *et al.*¹⁴).

It was encouraging to note that when administration of a systemic analgesic was reported, most papers specified a dose rate (Table 5) and reported dose rates were typically within the range recommended for rodents (see for example Flecknell *et al.*¹⁵). Buprenorphine was the most commonly used systemic analgesic in both time periods (Table 4) as had been previously reported in a survey of laboratory animal veterinarians¹⁶ and in an earlier literature review.⁶

Unfortunately, several of the findings from this survey suggest that analgesic administration in laboratory rodents is still not optimal. With the exception of combining a systemic analgesic with an anaesthetic regimen with an analgesic component, none of the papers included in the survey reported the use of administration of multimodal analgesia. Similarly, only two of the papers in the survey specified the administration of preoperative analgesia. Recommendations for improving the management of pain in rodents based on the existing literature are presented in Table 8.

Table 8 Recommendations for the alleviation of pain in laboratory rodents

Recommendation	Reference(s)
(1) Analgesic administration	
Administer at least one dose of systemic analgesia to all rodents undergoing recovery surgical procedures that are likely to be painful	Dobromylskyj <i>et al.</i> , ¹⁷ ACLAM ⁷
Consider the use of multimodal analgesia	
Consider the use of preoperative analgesia	
Match analgesic administration (dose rate, dose intervals and duration of administration) to the severity of the procedure	
(2) Reporting of analgesic administration and pain assessment	
Authors should include more information on analgesia and pain assessment in methods of peer-reviewed publications	Hawkins ⁴
In editorial policies/Instructions to Authors' editors should request that analgesic administration be specified and if analgesics were withheld to explain why	Hawkins, ⁴ Richardson and Flecknell ⁶
Authors of textbooks and training materials describing procedures commonly used for the production of transgenic mice (such as embryo transfer or vasectomy) should specify the administration of analgesic agents	
(3) Future research	
Further research into pain assessment in rodents and the dissemination of the findings of this research	Hawkins, ⁴ Flecknell and Roughtan, ¹⁸ Paul-Murphy <i>et al.</i> ¹⁹

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REFERENCES

- Russell WMS, Burch RL. *The Principles of Humane Experimental Technique*. London: Methuen, 1959
- House of Lords Select Committee on Animals in Scientific Procedures. *Report*. Vol. 1. London: HMSO, 2002
- Nuffield Council on Bioethics. *The Ethics of Research Involving Animals*. London: Nuffield Council on Bioethics, 2005
- Hawkins P. Recognising and assessing pain, suffering and distress in laboratory animals: a survey of current practices in the UK with recommendations. *Lab Anim* 2002;**36**:378–95. Full report: <http://www.lal.org.uk/pain/recognisingpain.pdf> (last checked 20 July 2007)
- Gauthier C. Overview and analysis of animal use in North America. *Altern Lab Anim* 2004;**32**(suppl. 1):275–85
- Richardson CA, Flecknell PA. Anaesthesia and post-operative analgesia following experimental surgery in laboratory rodents: are we making progress? *Altern Lab Anim* 2005;**33**:119–27
- American College of Laboratory Animal Medicine. Public statement: recommendations for the assessment and management of pain in rabbits and rodents. *J Am Assoc Lab Anim Sci* 2007;**46**:97–108
- Lamont LA, Mathews KA. Opioids, non-steroidal anti-inflammatories and analgesic adjuvants. In: Tranquilli WJ, Thurmon JC, Grimm KA, eds.

- Lumb and Jones' Veterinary Anesthesia and Analgesia*. 4th edn. Blackwell Publishing, 2007:241-72
- 9 Home Office. *Statistics of Scientific Procedures on Living Animals, Great Britain 2006*. London: HMSO, 2007
- 10 Malakoff D. The rise of the mouse, biomedicine's model mammal. *Science* 2001;**288**:248-53
- 11 Kohn DF, Clifford CB. Biology and diseases of rats. In: Fox JG, Anderson LC, Loew FM, Quimby FW, eds. *Laboratory Animal Medicine*. 2nd edn. San Diego: Academic Press, 2002:121-65
- 12 BVA/AFW/FRAME/RSPCA/UFAW Joint Working Group on Refinement. Refinement and reduction in the production of genetically modified mice. *Lab Anim* 2003;**37**(suppl. 1):S1-51
- 13 Wright-Williams SL, Courade J-P, Richardson CA, Roughan JV, Flecknell PA. Effects of vasectomy surgery and meloxicam treatment on faecal corticosterone and behaviour in two strains of laboratory mouse. *Pain* 2007;**130**:108-18
- 14 Carlsson H-E, Hagelin J, Hau J. Implementation of the 'Three Rs' in biomedical research. *Vet Rec* 2004;**154**:467-70
- 15 Flecknell PA, Richardson CA, Popovic A. Laboratory animals. In: Tranquilli WJ, Thurmon JC, Grimm KA, eds. *Lumb and Jones' Veterinary Anesthesia and Analgesia*. 4th edn. Blackwell Publishing, 2007:765-84
- 16 Hubbell JAE, Muir WW. Evaluation of a survey of the diplomates of the American College of Laboratory Animal Medicine on use of analgesic agents in animals used in biomedical research. *J Am Vet Med Assoc* 1996;**209**:918-21
- 17 Dobromylskyj P, Flecknell PA, Lascelles BD, Livingston A, Taylor P, Waterman-Pearson A. Management of postoperative and other acute pain. In: Flecknell PA, Waterman-Pearson A, eds. *Pain Management in Animals*. WB Saunders, 2000:81-147
- 18 Flecknell PA, Roughan JV. Assessing pain in animals - putting research into practice. *Anim Welf* 2004;**13**(suppl):S71-5
- 19 Paul-Murphy J, Ludders JW, Robertson SA, Gaynor JS, Hellyer PW, Wong PL. The need for a cross-species approach to the study of pain in animals. *JAVMA* 2004;**5**:692-7